

## 6. REFERÊNCIAS BIBLIOGRÁFICAS

AAS, E., BEYER, J., GOKSØYR, A. PAH in Fish Bile Detected by Fixed Wavelength Fluorescence. **Marine Environmental Research**, v. 46, n. 1-5, p. 225-228, 1998.

ALBERTS, B., JOHNSON, A., LEWIS, J., RAFF, M., ROBERTS, K. & WALTER, P. **Biologia Molecular da Célula**. 4. ed. Porto Alegre: Artmed, 1584 p, 2004.

ARIESE, F., BURGERS, I., OUDHOFF, K., RUTTEN, T., STROOMBERG, G., VETHAAK, D. **Comparison of Analytical Approaches for PAH Metabolites in Fish Bile Samples for Marine and Estuarine Monitoring**. Vrije Universiteit, Institute for Environmental Studies, Amsterdam, 29 p., 1997.

ARIESE, F., BEYER, J., JONSSON, G., VISA, C.P., KHRAN, M. **Review of analytical methods for determining metabolites of polycyclic aromatics compounds (PACs) in fish bile**. ICES Techniques in Marine Environmental Sciences, n. 39, 41 p., 2005b.

AZEVEDO, F.A. & CHASIN, A.A.M. **As bases toxicológicas da ecotoxicologia**. São Carlos: RiMa, 340 p., 2004.

BAMBER, S.D. & NAYLOR, E. Sites of release of Putative Sex Pheromone and Sexual Behaviour in Female *Carcinus maenas* (Crustacea: Decapoda). **Estuarine, Coastal and Shelf Science**, v. 44, p. 195–202, 1997.

BARRON, M.G., HEINTZ, R., RICE, S.D. Relative potency of PAHs and heterocycles as aryl hydrocarbon receptor agonist in fish. **Marine Environmental Research**, v. 58, p. 95-100, 2004.

BAUMARD, P., BUDZINSKI, H., GARRIGUES, P., SORBE, J. C., BURGEOT, T., BELLOCQ, J. Concentrations of PAHs (Polycyclic

Aromatic Hydrocarbons) in Various Marine Organisms in Relation to those in Sediments and to Trophic Level. **Marine Pollution Bulletin**, v. 36, p. 951-960, 1998.

BAUMARD, P., BUDZINSKI, H., GARRIGUES, P. PAHs in Arcachon Bay, France: Origin and Biomonitoring with Caged Organisms. **Marine Pollution Bulletin**, v. 36, p. 577-586, 1998.

BELLAS, J., THOR, P. Ecotoxicological evaluation of polycyclic aromatic hydrocarbons using marine invertebrate embryo-larval bioassays. **Marine pollution Bulletin**, v. 57, p. 493-502, 2008.

BOYSEN, G. & HECHT, S.S. Analysis of DNA and protein adducts of benzo[a]pyrene in human tissues using structure-specific methods. **Mutation Research**, v. 543, p. 17–30, 1997.

CAMUS, L., BIRKELY, S.R., JONES, M.B., BØRSETH, J.F., GRØSVIK, B.E., GULLIKSEN, B., LØNNE, O.J., REGOLI, F., DEPLEDGE, M.H. The Biomarker responses and PAH uptake in *Mya truncata* following exposure to oil-contaminated sediment in an Arctic fjord (Svalbard). **Science of the Total Environment**, v. 308, p. 221–234, 2003.

CASILLAS, R.P., CROW, S.A. Jr., HEINZE, T.M., CERNIGLIA, C.E. Initialoxidative and subsequent conjugatable metabolites produced during the metabolism of phenanthrene by fungi. **Journal of Industrial Microbiology**, v. 16, p. 205–215, 1996.

DAM, E., REWITZ, K.F., STYRISHAVE, B., ANDERSEN, O. Cytochrome P450 expression is moult stage specific and regulated by ecdysteroids and xenobiotics in the crab *Carcinus maenas*. **Biochemical and Biophysical Research Communications**, v. 377, p. 1135–1140, 2008.

DEPLEDGE, M.H., FOSSI, M. The role of biomarkers in environmental assessment in invertebrates. **Ecotoxicology**, v. 3, p. 161-172, 1994..

DISSANAYAKE, A., GALLOWAY, T.S., JONES, M.B. Physiological responses of juvenile and adult shore crabs *Carcinus maenas* (Crustacea:

Decapoda) to pyrene exposure. **Marine Environmental Research**, v. 66, p. 445–450, 2008.

DISSANAYAKE, A., GALLOWAY, T.S., JONES, M.B. Nutritional status of *Carcinus maenas* (Crustacea: Decapoda) influences susceptibility to contaminant exposure. **Aquatic Toxicology**, v. 89, p. 40–46, 2008.

DUNGAN, J.E., ICHIKAWA, G., STEPHENSON, M., CRANE, D.B., MCCALL, J., REGALADO, K. **Monitoring of coastal contaminants using sand crabs**. Relatório Final, Central Coast Regional Water Quality Control Board, Califórnia, 37p, 2005.

EICKHOFF, C.V., GOBAS, F.A.P.C., LAW, F.C.P. Screening pyrene metabolites in the hemolymph of dungeness crabs (*Cancer magister*) with Synchronous Fluorescence Spectrometry: Method development and application. **Journal of Environmental Toxicology and Chemistry**, v. 22, p. 59-66, 2003.

EURACHEM GUIDE, **The fitness for Purpose of analytical Methods**: a Laboratory Guide to Method Validation and Related Topics. Internet Version, 1998. Diponível em <http://www.eurachem.org/guidesanddocuments.htm>. Acesso em 20/12/2009.

ERUSTES, J.A., ANDRADE-EIROA, A., CLADERA, A., FORTEZA, R., CERDÀ, V. Fast sequential injection determination of benzo[A]pyrene using variable angle fluorescence with on-line solid-phase extraction. **Analytst**, v. 126, p. 451-456, 2001.

FARIAS, C. O., HAMACHER, C., WAGENER, A., SCOFIELD, A. Origin and degradation of hydrocarbons in mangrove sediments (Rio de Janeiro, Brazil) contaminated by an oil spill. **Organic Geochemistry**, v. 39, p. 289-307, 2008.

FERRARI, St., MANDEL, F., BERSET, J.D. Quantitative determination of 1-hydroxypyrene in bovine urine samples using high-performance liquid

chromatography with fluorescence and mass spectrometric detection. ***Chemosphere***, v. 47, p. 173–182, 2002.

FIGUEIREDO, L.H.M. **Investigação das contribuições orgânicas antrópicas e naturais em sedimentos costeiros utilizando-se hidrocarbonetos marcadores.** Tese de doutorado, Departamento de Química, PUC, Rio de Janeiro, 149 p., 1999.

FILLMANN, G., WATSON, G.M., FRANCIONI, E., READMAN, J.W., DEPLEDGE, M.H. A non-destructive assessment of the exposure of crabs to PAH using ELISA analyses of their urine and haemolymph. ***Marine Environmental Research***, v. 54, p. 823–828, 2002.

FOSSI, M.C., CASINI, S., SAVELLI, C., CORBELLINI, C., FRANCHI, E., MATEI, N., SANCHEZ-HERNANDEZ, J.C., CORSI, I., BAMBER, S., DEPLEDGE, M.H. Biomarker responses at different levels of biological organisation in crabs (*Carcinus aestuarii*) experimentally exposed to benzo(a)pyrene. ***Chemosphere***, v. 40, p. 861-874, 2000.

FRANCIONI, E., WAGENER, A. DE L.R., SCOFIELD, A. DE L., DEPLEDGE, M.H., CAVALIER, B. Biomonitoring of Polycyclic Aromatic Hydrocarbon in *Perna perna* from Guanabara Bay, Brazil. ***Environmental Forensics***, v. 6, p. 361–370, 2005.

FRANCIONI, E., WAGENER, A. DE L.R., SCOFIELD, A. DE L., DEPLEDGE, M.H., CAVALIER, B., SETTE, C.B., CARVALHOSA, L., LOZINSKY, C., MARIATH, R. Polycyclic aromatic hydrocarbon in intertidal mussel *Perna perna*: Space–time observations, source investigation and genotoxicity. ***Science of the Total Environment***, v. 372, p. 515–531, 2007a.

FRANCIONI, E., WAGENER, A. DE L.R., SCOFIELD, A. DE L., DEPLEDGE, M.H., CAVALIER, B. Evaluation of the mussel *Perna perna* as a biomonitor of polycyclic aromatic hydrocarbon (PAH) exposure and effects. ***Marine Pollution Bulletin***, v. 54, p. 329–338, 2007(b).

GRAVATO, C. & SANTOS, M.A. Juvenile sea bass liver biotransformation and erythrocytic genotoxic responses to pulp mill contaminants. ***Ecotoxicological Environmental and Safety***, v. 53, p. 104-112, 2002.

HOLTH, T.F., BEYLICH, B.A., SKARPHÉDINSDÓTTIR, H., LIEWENBORG, B., GRUNG, M., HYLLAND, K. Genotoxicity of environmentally relevant concentrations of water-soluble oil components in Cod (*Gadus morthua*). ***Environmental Science and Technology***, v. 43, p. 3329-3334, 2009.

HORNG, C.-Y., LIN, H.-C., LEE, W. A Reproductive Toxicology Study of Phenanthrene in Medaka (*Oryzias latipes*). ***Archive of Environmental Contaminants and Toxicology***, v. 58, p. 131–139, 2010.

IKENAKA, Y., ISHIZAKA, M., EUN, H., MIYABARA, Y., 2007. Glucose-sulfate conjugates as a new phase II metabolite formed by aquatic crustaceans. ***Biochemical and Biophysical Research Communications***, v. 360, p. 490–495, 2007.

IRWIN, R.J., MOUWERIK, M.V., STEVENS, L., STEVENS, M.D., BASHAM, W. ***Environmental Contaminants Encyclopedia Phenanthrene Entry***. National Park Service Water Resources Divisions, Water Operations Branch, Colorado, 57 p., 1997.

JACOB, J., RAAB, G., SOBALLA, V., SCHMALIX, W.A., GRIMMER, G., GREIM, H., DOEHMER, J., SEIDEL, A. Cytochrome P450-mediated activation of phenanthrene in genetically engineered V79 Chinese hamster cells. ***Environmental Toxicology and Pharmacology***, v. 1, p. 1-11, 1996.

JOHNSEN, A.R., WICK, L.Y., HARMS, H. Principles of microbial PAH-degradation in soil. ***Environmental Pollution***, v. 113, p. 71-84, 2005.

KOENIG, S., SVAGE, C., KIM, J.P. Non-destructive assessment of polycyclic aromatic hydrocarbon (PAH) exposure by fluorimetric analysis of crab urine. ***Marine Pollution Bulletin***, v. 56, p. 2003–2008, 2008.

LIMA, E.F.A. **Acumulação de hidrocarbonetos policíclicos aromáticos e metais traço em invertebrados marinhos e avaliação do uso de biomarcadores celulares e bioquímicos no biomonitoramento.** Tese de doutorado, Departamento de Química, PUC, Rio de Janeiro, 167p., 2001.

LIPIATOU, E., TOLOSA, I., SIMÓ, R., BOULOUBASSI, I., DACHS, J., MARTI, S., SICRE, M.-A., BAYONA, J.M., GRIMALT, J.O., SALIOT, A., ALBAIGÉS, J. Mass budget and dynamics of polycyclic aromatic hydrocarbons in the mediterranean sea. **Deep-Sea Research II**, v. 44, p. 881-905, 1997.

LISOWSKA, K., DŁUGOŃSKI, J., FREEMAN, J.P., CERNIGLIA, C.L. The effect of the corticosteroid hormone cortexolone on the metabolites produced during phenanthrene biotransformation in *Cunninghamella elegans*. **Chemosphere**, v.64, p. 1499-1506, 2006..

LUO, X.J., CHEN, S.J., MAI, B.X., YANG, Q.S., SHENG, G.Y., FU, J.M. Polycyclic aromatic hydrocarbons in suspended particulate matter and sediments from the Pearl River Estuary and adjacent coastal areas, China. **Environmental Pollution**, v. 139, p. 9-20, 2006.

LUTHE, G., STROOMBERG, G.J., ARIESE, F., BRINKMAN, U.A.TH., VAN STRAALEN, N.M., 2002. Metabolism of 1-fluoropyrene and pyrene in marine flatfish and terrestrial isopods. **Environmental Toxicology and Pharmacology**, 12, 221- 229.

MENICONI, M.F.G.; GABARDO, I.T.; CARNEIRO, M.E.R.; BARBANTI, S.M. ; SILVA, G.C. ; MASSONE, C.G. 2002; Brazilian Oil Spills Chemical Characterization – Case Studies. **Environmental Forensics**, 3, 303-321.

MENICONI, M.F.G.. **Hidrocarbonetos policíclicos aromáticos no meio ambiente:** diferenciação de fontes em sedimentos e metabólitos em bile de peixes. Tese de Doutorado, Centro de Ciências Exatas e da Terra, UFRN, Natal, 213 p., 2007.

MORALES-CASELLES, C. MARTÍN-DÍAZ, M.L., RIBA, I., SARASQUETE, C., DELVALLS, T.A. Sublethal responses in caged organisms exposed to sediments affected by oil spills. **Chemosphere**, v. 72, p. 819-825, 2008.

MORRISON, R. D. **Environmental forensics: principles and applications**. Boca Raton: CRC Press LLC, 1999. (Digital)

NASCIMENTO, S.A. **Biologia do caranguejo-uçá (*Ucides cordatus*)**. Adema, Aracajú, v. 1, 48 p., 1993

NEFF, M., 1979. **Polycyclic aromatic hydrocarbons in the aquatic environment : sources, fates and biological effects**. London: Applied Science Publishers.

NEFF, M. **Bioaccumulation in marine organisms: effect of contaminants from oil well produced water**. Coastal Resources and Environmental Management. Duxbury, Massachusetts, 453 p., 2002.

NETTO, A.D.P., MOREIRA, J.C., DIAS, A.E.X.O., ARBILLA, G., FERREIRA, L.F.V., OLIVEIRA, A.S., BAREK, J., 2000. Avaliação da contaminação humana por hidrocarbonetos policíclicos aromáticos (HPAs) e seus derivados nitrados (NHPAs): uma revisão metodológica. **Química Nova**, v. 23, p. 765-773.

NEWMAN, M.C., ROBERTS, M.H., HALE, R.C., **Coastal and estuarine risk assessment**. Florida: Lewis Publishers, 2002. (Digital)

NUDI, A.H. **Avaliação da contaminação de manguezais da Baía de Guanabara utilizando caranguejos *Ucides cordatus* como bioindicador de poluentes de petróleo e desenvolvimento de metodologias de análise**. Tese de doutorado, Departamento de Química, PUC, Rio de Janeiro, 233p., 2005

NUDI, A.H., WAGENER, A.L.R., FRANCIONE, E., SCOFIELD, A.L., SETTE, C.B., VEIGA, A. Validation of *Ucides cordatus* as a bioindicator of oil contamination and bioavailability in mangroves by evaluating sediment

and crab PAH records. **Environment International**, v. 33, p. 315-327, 2007.

NUDI, A.H., WAGENER, A.L.R, FRANCIONE, E., SETTE, C.B., SARTORI, A. V., SCOFIELD, A.L. Biomarkers of PAHs exposure in crabs *Ucides cordatus*: Laboratory assay and field study. **Environment Research**, v. 110, p. 137-145, 2010.

O'CONNOC & SQUIBB, 1989 *apud* NEFF, M. **Bioaccumulation in marine organisms: effect of contaminants from oil well produced water.** Coastal Resources and Environmental Management, Massachusetts: Duxbury, 2002.

OLIVEIRA, M., PACHECO, M., SANTOS, M.A. Cytochrome P4501A, genotoxic and stress responses in golden grey mullet (*Liza aurata*) following short-term exposure to phenanthrene. **Chemosphere**, v. 66, p. 1284-1291, 2007.

ONYEMAUWA, F., RAPPAPORT, S.M., SOBUS, J.R., GAJDOŠOVÁ, D., WU, R., WAIDYANATHA, S. Using liquid chromatography–tandem mass spectrometry to quantify monohydroxylated metabolites of polycyclic aromatic hydrocarbons in urine. **Journal of Chromatography B**, v.877, p. 1117-1125, 2009.

ORBEA, A., ORTIZ-ZARRAGOITIA, M., SOLE, M., PORTE, C., CAJARAVILLE, M.P. Antioxidant enzymes and peroxisome proliferation in relation to contaminant body burdens of PAHs and PCBs in bivalve molluscs, crabs and fish from the Urdaibai and Plentzia estuaries (Bay of Biscay). **Aquatic Toxicology**, v. 58, p. 75-98, 2002.

PÉREZ, S. & BARCELÓ, D. Determination of Polycyclic Aromatic Hydrocarbons in Sewage Reference Sludge by Liquid Chromatography-Atmospheric-Pressure Chemical-Ionization Mass Spectrometry. **Chromatographia**, v. 53, p. 475-480, 2001

PRUELL, R.J.; *et al*, 1989. **Contaminant Concentrations in Sediments from the Foul Area Disposal Site in Massachusetts Bay.** U.S. Environmental Protection Agency and Science Applications International Corporation (SAIC), 25 p., 1989.

RAPPAPORT, S.M., WAIDYANATHA, S., YEOWELL-O'CONNELL, K., ROTHMAN, N., SMITH, M.T., ZHANG, L., QU, Q., SHORE, R., LI, G., YIN, S. Protein adducts as biomarkers of human benzene metabolism. **Chemical-Biological Interactions**, v. 153-154, p. 103-109, 2005.

RISER-ROBERTS. **Remediation of petroleum contaminated soil: biological, physical, and chemical processes.** Florida: Lewis Publishers, 1998. (Digital)

RIZEL, P. **Estudos qualitativos para determinação de metabólitos de pireno empregando CG/EM.** Rio de Janeiro, PUC, Departamento de Química - Tese de doutorado 53 p., 2003.

RUPPERT, E.E. & BARNES, R.D., 1996. **Zoologia dos invertebrados.** 6. Ed. São Paulo: Roca, 1029p.

SHEN, R.-Y., LUO, Y.-M., FENG, S., ZHANG, G.-Y., WU, L.-H., LI, Z.-G., TENG, Y., CHRISTIE, P. Benzo[a]pyrene and Phenanthrene in Municipal Sludge from the Yangtze River Delta, China. **Pedosphere**, v. 19, p. 523–531, 2009.

SMITH, D.R., ROBB, D.B., BLADES, M.W. Comparison of Dopants for Charge Exchange Ionization of Nonpolar Polycyclic Aromatic Hydrocarbons with Reversed-Phase LC-APPI-MS. **Journal of American Society of Mass Spectrometry**, v. 20, p. 73–79, 2009.

STAAL, Y.C.M., HEBELS, D., van HERWILNEN, M.H.M., GOTTSCHALK, R.W.H., van SCHOOTEN, F.J., van DELFT, J.H.M. Binary PAH mixtures cause additive or antagonistic effect on gene expression but synergic effect on DNA adduct formation. **Carcinogenesis**, v. 28, p. 2632-2640, 2007.

STROOMBERG, G.J. Pyrene metabolites in isopods (Crustácea) as biomarker for PAH exposure in terrestrial ecosystems. Tese de doutorado, Institute of Ecological Science, Vrije Universiteit, Amsterdam, 176p., 2002.

STROOMBERG,,G.J., ZAPPEY, H., STEEN, R.J.C.A., VAN GESTEL, C.A.M., ARIESE, F., VELTHORST, N.H., VAN STRAALEN, N.M. PAH biotransformation in terrestrial invertebrates—a new phase II metabolite in isopods and springtails. **Comparative Biochemistry and Physiology, Part C**, v. 138, p. 129–137, 2004.

SUN, Y., YU, H., ZHANG, J., YIN, Y., SHI, H., WANG, X. Bioaccumulation, depuration and oxidative stress in fish *Carassius auratus* under phenanthrene exposure. **Chemosphere**, v. 63, p. 1319-1327, 2006.

UMBUZEIRO, G.A., KUMMROW, F., ROUBICEK, D.A., TOMINANGA, M.Y. Evaluation of the water genotoxicity from Santos estuary (Brazil) in relation to the sediment contamination and effluent discharges. **Environmental International**, v. 32, p. 359-364, 2006.

VALERO-NAVARRO, A., FERNÁNDEZ-SÁNCHEZ, J.F., MEDINA-CASTILLO, A.L., FERNANDÉZ-IBÁÑEZ, F., SEGURA-CARRETO, A., IBÁÑEZ, J.M., FERNÁNDEZ-GUTIÉRREZ, A. A rapid, sensitive screening test for polycyclic aromatic hydrocarbons applied to Antarctic water. **Chemosphere**, v. 67, p. 903-910, 2007.

VUORINEN, P.J., KEINÄNEN, M., VUONTISJÄRVI, H., BARSIENE, J., BROEG, K., FÖRLIN, L., GERCKEN, J., KOPECKA, J., KÖHLER, A., PARKKONEN, J., PEMPKOWIAK, J., SCHIEDEK, D. Use of biliary PAH metabolites as a biomarker of pollution in fish from the Baltic Sea. **Marine Pollution Bulletin**, v. 53, p. 479-487, 2006.

WATSON, G.M., ANDERSEN, O.-K., GALLOWAY, T.S., DEPLEDGE, M.H. Rapid assessment of polycyclic aromatic hydrocarbon (PAH) exposure in decapod crustaceans by fluorimetric analysis of urine and haemolymph. **Aquatic Toxicology**, v. 67, p. 127–142, 2004.

WATSON, G.M., ANDERSEN, O.-K., DEPLEDGE, M.H., GALLOWAY, T.S. Detecting a field gradient of PAH exposure in decapod crustacea using a novel urinary biomarker. **Marine Environmental Research**, v. 58, p. 257–261, 2004(b).

WU, R., WAIDYANATHA, S., HENDERSON, A.P., SERDAR, B., ZHENG, Y., RAPPAPORT, S.M. Determination of dihydroxynaphthalenes in human urine by gas chromatography-mass spectrometry. **Journal Chromatography B**, v. 877, p. 206-213, 2005.

YIN, Y, JIA, H., SUN, Y, YU, H., WANG, X., WU, J., XUE, Y. Bioaccumulation and ROS generation in liver of *Carassius auratus*, exposed to phenanthrene. **CBP**, v. 145, p. 288-293, 2007.

YUNKER, M.B., CRETNEY, W.J., IKONOMOU, M.G. Assessment of Chlorinated Dibenzo-*p*-dioxin and Dibenzofuran Trends in Sediment and Crab Hepatopancreas from Pulp Mill and Harbor Sites Using Multivariate- and Index-Based Approaches. **Environmental Science & Technology**, v. 36, p. 1869-1978. 2002.

ZEMANEK, M.G., POLLARD, S.J.T., KENEFICK, S.L., HRUDEY, S.E. Multi-phase portioning and co-solvent effects for polynuclear aromatic hydrocarbons (PAH) in authentic petroleum – and creosote-contaminated soils. **Environmental Pollution**, v. 98, p. 239-252, 1997.

ZHANG, J., CAI, L., YUAN, D., CHEN, M. Distribution and sources of polynuclear aromatic hydrocarbons in Mangrove surficial sediments of Deep Bay, China. **Marine Pollution Bulletin**, v. 49, p. 479–486, 2004.

ZHU, S., LI, L., THORNTON, C., CARVALHO, P., AVERY, B.A., WILLETT, K.L. Simultaneous determination of benzo(a)pyrene and eight of its metabolites in *Fundulus heteroclitus* bile using ultra-performance liquid chromatography with mass spectrometry. **Journal of Chromatography B**, 863, 141-149, 2008.

## 7. ANEXOS

Anexo 1. Áreas dos picos de possíveis metabólitos por CLAE/EM nas amostras controle

Picos/Amostras	Controle 48h	Controle 48h hidrolisada	Controle 72h	Controle 72h hidrolisada
<b>Fenantreno diol</b>	nd	nd	nd	nd
<b>Fenantreno epóxido</b>	nd	256524	nd	213274
<b>Trihidroxi fenantreno</b>	nd	nd	nd	nd
<b>Não identificado 1</b>	nd	nd	nd	nd
<b>Não identificado 2</b>	nd	nd	nd	nd
<b>Fenantreno monolepoxido</b>	nd	397932	nd	307199
<b>Fenantreno diolepoxido</b>	nd	139105	nd	147185
<b>Fenantreno glicosideo</b>	nd	nd	nd	161284
<b>Fenantreno tetrol</b>	nd	nd	nd	nd
<b>Não identificado 3</b>	nd	nd	nd	nd
<b>Fenantreno ortoquinona</b>	nd	nd	nd	91156
<b>Monohidroxi fenantreno</b>	nd	nd	nd	nd
<b>Não identificado 3</b>	nd	nd	nd	nd
a	17716	nd	16258	nd
b	159343	nd	196299	nd
c	103560	nd	114409	nd
d	165894	nd	154613	nd
w	nd	nd	nd	nd
Y	nd	nd	nd	nd
Z	nd	nd	nd	nd

Anexo 2. Áreas dos picos de possíveis metabólitos por CLAE/EM nas amostras inoculadas com fenantreno

Picos/Amostras	Fenantreno 24h	Fenantreno 24h hidrolisada	Fenantreno 48h	Fenantreno 48 hidrolisada
<b>Fenantrneo diol</b>	nd	nd	nd	nd
<b>Fenantreno epoxido</b>	nd	204649	nd	181217
<b>Trihidroxi fenantreno</b>	nd	nd	nd	nd
<b>Não identificado 1</b>	nd	nd	nd	nd
<b>Não identificado 2</b>	nd	nd	nd	nd
<b>Fenantreno monolepoxido</b>	nd	288572	nd	289902
<b>Fenantreno diolepoxido</b>	nd	119323	nd	133533
<b>Fenantrneo glicosideo</b>	nd	nd	nd	nd
<b>Fenantreno tetrol</b>	nd	nd	nd	nd
<b>Não identificado 3</b>	nd	nd	nd	nd
<b>Fenantreno ortoquinona</b>	nd	nd	221006	nd
<b>Monohidroxi fenantreno</b>	nd	nd	nd	nd
<b>Não identificado 3</b>	nd	nd	nd	nd
a	17488	nd	76143	nd
b	86673	nd	243253	nd
c	40895	nd	nd	nd
d	101752	nd	291545	nd
w	nd	nd	nd	nd
Y	nd	nd	56566	nd
Z	nd	nd	72506	nd

Anexo 3. Áreas dos picos de possíveis metabólitos por CLAE/EM nas amostras inoculadas com 1-Metil fenantreno

Picos/Amostras	1-Metil fenantrneo 24h	1-Metil fenantrneo 24h hidrolisada	1-Metil fenantreno 48h	1-Metil fenantreno 48h hidrolisada
<b>Fenantrneo diol</b>	nd	nd	nd	nd
<b>Fenantreno epóxido</b>	nd	246895	nd	250999
<b>Trihidroxi fenantreno</b>	nd	nd	nd	nd
<b>Não identificado 1</b>	nd	nd	nd	nd
<b>Não identificado 2</b>	nd	nd	nd	nd
<b>Fenantreno monolepoxido</b>	nd	335928	nd	394381
<b>Fenantreno diolepoxido</b>	nd	141497	nd	133055
<b>Fenantrneo glicosideo</b>	nd	58811	nd	nd
<b>Fenantreno tetrol</b>	nd	nd	nd	79494
<b>Não identificado 3</b>	nd	nd	nd	nd
<b>Fenantreno ortoquinona</b>	nd	130567	nd	148400
<b>Monohidroxi fenantreno</b>	nd	nd	nd	nd
<b>Não identificado 3</b>	nd	n.f	nd	96596
a	58630	nd	nd	nd
b	201145	nd	nd	nd
c	434218	nd	nd	nd
d	nd	nd	nd	nd
w	nd	nd	5142232	nd
Y	nd	nd	nd	nd
Z	nd	nd	nd	nd

Anexo 4. Áreas dos picos de possíveis metabólitos por CLAE/EM nas amostras inoculadas com 2,6,9-Trimetil fenantreno

Picos/Amostras	2,6,9-Trimetil fenantreno 24h	2,6,9-Trimetil fenantreno 24h hidrolisada	2,6,9-Trimetil fenantreno 48h	2,6,9-Trimetil fenantreno 48h hidrolisada
<b>Phenanthrene Diol</b>	nd	nd	nd	nd
<b>Phenanthrene epoxide</b>	nd	211474	nd	163993
<b>Trihydroxyphenanthrene</b>	nd	nd	nd	nd
<b>Não identificado 1</b>	nd	nd	nd	nd
<b>Não identificado 2</b>	nd	nd	nd	nd
<b>Phenanthrene monolepoxide</b>	nd	303633	nd	296263
<b>Phenanthrene diolepoxide</b>	nd	102758	nd	172322
<b>Phenanthrene glicoside</b>	nd	nd	nd	nd
<b>Phenanthrene tetrol</b>	nd	nd	nd	nd
<b>Não identificado 3</b>	113820	nd	<b>291773*</b>	nd
<b>Phenanthrene orthoquinone</b>	nd	nd	<b>291773*</b>	nd
<b>OH-phenanthrene</b>	nd	nd	nd	nd
<b>Não identificado 4</b>	nd	nd	nd	nd
a	38190	nd	43269	nd
b	497690	nd	202469	nd
c	325295	nd	335832	nd
d	nd	nd	nd	nd
w	nd	nd	nd	nd
Y	nd	nd	nd	nd
Z	nd	nd	nd	nd

Anexo 5. Resultado contagem de células do ensaio do MN no bioensaio 2 dos organismos expostos ao sedimento

<b>Ensaio/Tempo</b>	<b>Células normais</b>	<b>1MN</b>	<b>2 ou +MN</b>	<b>MN total</b>
Sed/24h	1360	1	0	1
Sed/24h	1339	2	0	2
Sed/24h	1349	2	0	2
Sed/24h	1928	0	0	0
Sed/24h	448	0	0	0
Sed/24h	1026	2	0	2
Sed/48h	1786	2	0	2
Sed/48h	2625	2	1	3
Sed/48h	3197	0	1	1
Sed/72h	2042	0	0	0
Sed/72h	2225	4	0	4
Sed/72h	1626	6	0	6
Sed/96h	1347	3	0	3
Sed/96h	865	5	0	5

Anexo 6. Resultado contagem de células do ensaio do MN no bioensaio 3

<b>Tempo</b>	<b>Ensaio</b>	<b>Ensaio/Tempo</b>	<b>Células normais</b>	<b>1MN</b>	<b>2 ou +MN</b>
24h	Controle	Cont. 24h	716	0	0
72h	Controle	Cont. 72h	1354	0	0
24h	Fen	Fen 24h	762	0	0
24h	Fen	Fen 24h	611	0	0
24h	Fen	Fen 24h	x	0	0
24h	Fen	Fen 24h	332	0	0
24h	Fen	Fen 24h	x	0	0
48h	Fen	Fen 48h	362	0	
48h	Fen	Fen 48h	1501	0	5
72h	Fen	Fen 72h	x	0	0
24h	TriMeFen	TriMeF 24h	1816	1	1
24h	TriMeFen	TriMeF 24h	1091	0	0
24h	TriMeFen	TriMeF 24h	1127	0	1
24h	TriMeFen	TriMeF 24h	1091	0	0
48h	TriMeFen	TriMeF 48h	x	0	0
48h	TriMeFen	TriMeF 48h	300	0	0
72h	TriMeFen	TriMeF 72h	x	0	0

Anexo 7. Resultado contagem de células do ensaio do MN no bioensaio 3 (cont.)

<b>Tempo</b>	<b>Ensaio</b>	<b>Ensaio/Tempo</b>	<b>Células normais</b>	<b>1MN</b>	<b>2 ou +MN</b>
24h	1-MeFen	1-MeF 24h	596	0	0
24h	1-MeFen	1-MeF 24h	x	0	0
24h	1-MeFen	1-MeF 24h	816	0	0
24h	1-MeFen	1-MeF 24h	1004	1	2
24h	1-MeFen	1-MeF 24h	847	0	1
48h	1-MeFen	1-MeF 48h	178	0	0
48h	1-MeFen	1-MeF 48h	1088	1	0
48h	1-MeFen	1-MeF 48h	965	0	0
48h	1-MeFen	1-MeF 48h	938	2	0
72h	1-MeFen	1-MeF 72h	x	0	0
72h	1-MeFen	1-MeF 72h	x	0	0

Anexo 8. Concentração dos hidrocarbonetos policíclicos aromáticos individuais em amostras de hepatopâncreas do bioensaio 1 (ng.g<sup>-1</sup>)

Amostra	Maio 09 Fen 12h	Maio 089 Fen 24h	Maio 09 Fen 48h	Maio 09 Fen 72h
NAF	19,77	27,78	21,13	<LD
1ME_NAF	11,92	11,12	7,73	<LD
2Me_NAF	11,33	17,54	13,28	<LD
C <sub>2</sub> _NAF	52,03	61,98	37,54	N/F
C <sub>3</sub> _NAF	29,29	17,06	31,32	5,22
C <sub>4</sub> _NAF	237,49	-13,29	77,84	17,71
ACENAFTY	<LD	<LD	<LD	nd
ACE	<LD	<LD	<LD	<LD
FLUOR	34,43	47,31	45,90	<LQ
C <sub>1</sub> _FLUOR	56,34	106,72	75,12	11,31
C <sub>2</sub> _FLUOR	381,97	-9,21	509,29	85,89
C <sub>3</sub> _FLUOR	481,66	1201,76	642,22	175,87
DBZTIOF	177,77	466,57	115,55	70,13
C <sub>1</sub> _DBZTIOF	20,20	1,38	22,69	8,44
C <sub>2</sub> _DBZTIOF	33,40	82,04	37,54	18,29
C <sub>3</sub> _DBZTIOF	52,95	124,71	53,60	28,76
FEN	16047,17	50637,03	21396,22	7987,62
C <sub>1</sub> _FEN	37,48	-11,22	49,97	19,37
C <sub>2</sub> _FEN	241,33	-11,22	321,77	114,87
C <sub>3</sub> _FEN	945,06	-4,26	1260,07	301,71
C <sub>4</sub> _FEN	119,83	240,15	159,77	45,16
ANT	128,80	503,08	171,73	58,80
FLUORAN	19,50	18,36	26,00	8,64
PIR	43,45	17,28	57,94	18,56
C <sub>1</sub> _PIR	84,85	34,47	113,13	35,35
C <sub>2</sub> _PIR	24,73	1,57	32,97	13,26
Bz(a)ANT	16,43	<LD	21,91	<LD
CRIS	2,32	<LD	<LD	7,51
C <sub>1</sub> _CRIS	<LD	<LD	<LD	<LD
C <sub>2</sub> _CRIS	<LD	<LD	<LD	<LD
Bz(b)FLUOR	<LD	<LD	<LD	<LD
Bz(k)FLUOR	6,76	<LD	<LQ	<LD
Bz(e)PIR	<LQ	4,96	<LQ	N/F
Bz(a)PIR	nd	<LD	<LD	<LD
Perileno	nd	2,14	nd	nd
Indeno(123-cd)PIR	<LD	<LD	<LD	<LD
DBz(ah)ANT	<LD	<LD	<LD	nd
Bz(ghi)PERIL	<LD	<LD	<LD	<LD
p-TERPhd14	117,83	98,34	124,20	83,56
16 HPA	16318,63	51250,83	21740,83	8081,14
Σ HPA	19318,24	53575,80	25302,25	9032,47

Anexo 9. Concentração dos hidrocarbonetos policíclicos aromáticos individuais em amostras de hepatopâncreas do bioensaio 1 (ng.g<sup>-1</sup>)

<b>Amostra</b>	<b>Maio 09 Óleo 12h</b>	<b>Maio 09 Óleo 24h</b>	<b>Maio 09 Óleo 48h</b>	<b>Maio 09 Óleo 72h</b>
NAF	13,26	31,50	22,42	39,54
1ME_NAF	124,35	290,01	376,66	342,59
2Me_NAF	156,77	322,19	410,88	422,73
C2_NAF	792,04	2795,62	3702,55	2698,31
C3_NAF	423,85	1222,89	1503,39	973,70
C4_NAF	388,28	878,91	989,51	644,68
ACENAFTY	nd	10,08	11,88	10,39
ACE	21,52	71,78	80,37	59,94
FLUOR	27,28	82,49	84,27	73,23
C1_FLUOR	135,85	225,98	227,34	208,65
C2_FLUOR	404,24	344,16	417,53	<LD
C3_FLUOR	317,43	233,23	263,51	265,37
DBZTIOF	26,23	70,41	61,66	53,81
C1_DBZTIOF	56,53	113,53	138,02	0,65
C2_DBZTIOF	50,74	82,28	125,23	86,74
C3_DBZTIOF	32,72	48,07	75,27	92,03
FEN	144,06	434,44	361,34	261,46
C1_FEN	261,10	453,77	552,20	388,77
C2_FEN	244,89	303,92	417,64	276,63
C3_FEN	387,37	14,66	146,60	11,57
C4_FEN	30,67	23,74	22,71	57,40
ANT	10,88	68,40	71,28	60,47
FLUORAN	8,64	10,69	15,49	22,52
PIR	27,26	18,29	28,66	28,73
C1_PIR	19,02	6,24	10,81	12,33
C2_PIR	15,07	0,85	3,93	12,17
Bz(a)ANT	6,68	<LD	0,88	19,92
CRIS	3,28	<LD	4,40	6,17
C1_CRIS	<LD	<LD	<LD	<LD
C2_CRIS	<LD	<LD	<LD	<LD
Bz(b)FLUOR	nd	<LD	<LD	<LD
Bz(k)FLUOR	<LD	<LD	<LD	<LD
Bz(e)PIR	nd	2,85	3,07	2,13
Bz(a)PIR	<LD	<LD	<LD	<LD
Perileno	nd	1,19	1,36	1,01
Indeno(123-cd)PIR	<LD	<LD	<LD	<LD
DBz(ah)ANT	<LD	<LD	<LD	<LD
Bz(ghi)PERIL	<LD	<LD	<LD	<LD
p-TERPhd14	86,40	76,45	75,70	88,97
16 HPA	262,87	727,67	680,98	582,36
$\Sigma$ HPA	4130,04	8162,16	10130,86	7133,65

Anexo 10. Concentração dos hidrocarbonetos policíclicos aromáticos individuais em amostras de hepatopâncreas do bioensaio 1 (ng.g<sup>-1</sup>)

Amostra	Ago 09 Campo	Ago 09 To	Ago 09 Cont. 24hs	Ago 09 Cont. 48hs	Ago 09 Cont. 48hs
NAF	27,10	nd	38,46	34,24	nd
1ME_NAF	19,08	nd	15,67	32,14	nd
2Me_NAF	34,67	nd	21,41	26,13	nd
C <sub>2</sub> _NAF	nd	nd	nd	nd	nd
C <sub>3</sub> _NAF	<LD	<LD	<LD	<LD	<LD
C <sub>4</sub> _NAF	<LD	<LD	<LD	<LD	<LD
ACENAFTY	19,31	nd	15,65	nd	nd
ACE	nd	nd	nd	nd	nd
FLUOR	nd	nd	20,19	nd	nd
C <sub>1</sub> _FLUOR	<LD	<LD	<LD	<LD	<LD
C <sub>2</sub> _FLUOR	<LD	<LD	<LD	<LD	<LD
C <sub>3</sub> _FLUOR	<LD	<LD	<LD	<LD	<LD
DBZTIOF	nd	nd	<LQ	nd	nd
C <sub>1</sub> _DBZTIOF	<LD	<LD	<LD	<LD	<LD
C <sub>2</sub> _DBZTIOF	<LD	<LD	<LD	<LD	<LD
C <sub>3</sub> _DBZTIOF	<LD	<LD	<LD	<LD	<LD
FEN	116,46	nd	46,95	31,39	37,52
C <sub>1</sub> _FEN	<LD	<LD	<LD	<LD	<LD
C <sub>2</sub> _FEN	<LD	<LD	<LD	<LD	<LD
C <sub>3</sub> _FEN	<LD	<LD	<LD	<LD	<LD
C <sub>4</sub> _FEN	<LD	<LD	<LD	<LD	<LD
ANT	nd	nd	nd	nd	nd
FLUORAN	nd	nd	nd	nd	nd
PIR	nd	nd	nd	nd	nd
C <sub>1</sub> _PIR	<LD	<LD	<LD	<LD	<LD
C <sub>2</sub> _PIR	<LD	<LD	<LD	<LD	<LD
Bz(a)ANT	8,15	nd	nd	nd	nd
CRIS	<LQ	nd	nd	nd	nd
C <sub>1</sub> _CRIS	<LD	<LD	<LD	<LD	<LD
C <sub>2</sub> _CRIS	<LD	<LD	<LD	<LD	<LD
Bz(b)FLUOR	nd	nd	nd	nd	nd
Bz(k)FLUOR	nd	nd	nd	nd	nd
Bz(e)PIR	nd	nd	nd	nd	nd
Bz(a)PIR	nd	nd	nd	nd	nd
Perileno	nd	nd	nd	nd	nd
Indeno(123- cd)PIR	nd	nd	nd	nd	nd
DBz(ah)ANT	nd	nd	nd	nd	nd
Bz(ghi)PERIL	nd	nd	nd	nd	nd
p-TERPhd14	76,28	83,06	71,71	66,73	65,18
16 HPA	171,01	0,00	121,25	65,63	37,52
Σ HPA	224,76	0,00	158,34	123,90	37,52

Anexo 11 Concentração dos hidrocarbonetos policíclicos aromáticos individuais em amostras de hepatopâncreas do bioensaio 1 (ng.g-1)

<b>Amostra</b>	<b>Ago 09 Óleo 24hs</b>	<b>Ago 09 Óleo 48hs</b>	<b>Ago 09 Oleo 48hs</b>
NAF	144,30	46,34	39,69
1ME_NAF	366,75	150,17	164,54
2Me_NAF	490,36	179,95	214,84
C2_NAF	8056,10	1897,76	2026,68
C3_NAF	22778,71	1671,33	1530,64
C4_NAF	9863,39	1053,27	1091,90
ACENAFTY	23,75	18,12	31,98
ACE	37,06	44,12	65,78
FLUOR	76,27	55,77	104,22
C1_FLUOR	170,63	<LD	174,67
C2_FLUOR	<LD	<LD	<LD
C3_FLUOR	<LD	<LD	<LD
DBZTIOF	43,80	nd	46,06
C1_DBZTIOF	<LD	<LD	<LD
C2_DBZTIOF	<LD	<LD	<LD
C3_DBZTIOF	<LD	<LD	<LD
FEN	233,60	200,66	232,63
C1_FEN	3921,28	18504,79	4603,71
C2_FEN	<LD	<LD	<LD
C3_FEN	<LD	<LD	<LD
C4_FEN	<LD	<LD	<LD
ANT	75,42	109,30	165,49
FLUORAN	nd	nd	nd
PIR	nd	nd	nd
C1_PIR	<LD	<LD	<LD
C2_PIR	<LD	<LD	<LD
Bz(a)ANT	nd	nd	nd
CRIS	nd	nd	nd
C1_CRIS	<LD	<LD	<LD
C2_CRIS	<LD	<LD	<LD
Bz(b)FLUOR	nd	nd	nd
Bz(k)FLUOR	nd	nd	nd
Bz(e)PIR	nd	nd	nd
Bz(a)PIR	nd	nd	nd
Perileno	nd	nd	nd
Indeno(123-cd)PIR	nd	nd	nd
DBz(ah)ANT	nd	nd	nd
Bz(ghi)PERIL	nd	nd	nd
p-TERPHd14	62,43	73,02	60,40
16 HPA	590,39	474,31	639,78
$\Sigma$ HPA	46281,41	23931,58	10492,83

Anexo 12 Concentração dos hidrocarbonetos policíclicos aromáticos individuais no óleo árabe leve e concentração real inoculada no bioensaio 1 (ng.g<sup>-1</sup>)

Amostra	óleo árabe	Conc. Inoculada
NAF	1452,1	2904,2
1ME_NAF	18852,1	37704,2
2Me_NAF	10558,1	21116,2
C2_NAF	61032,3	122064,6
C3_NAF	55181,6	110363,2
C4_NAF	28911,0	57822
ACE	469,6	939,2
ACENAFTY	1885,6	3771,2
FLUOR	1404,5	2809
C1_FLUOR	3724,8	7449,6
C2_FLUOR	5529,5	11059
C3_FLUOR	3934,1	7868,2
DBZTIOF	804,7	1609,4
C1_DBZTIOF	1564,3	3128,6
C2_DBZTIOF	895,2	1790,4
C3_DBZTIOF	364,8	729,6
FEN	4914,4	9828,8
C1_FEN	8051,6	16103,2
C2_FEN	4921,2	9842,4
C3_FEN	1272,9	2545,8
C4_FEN	259,8	519,6
ANT	1033	2066
FLUORAN	27,2	54,4
PIR	125,1	250,2
C1_PIR	84	168
C2_PIR	23	46
Bz(a)ANT	1,1	2,2
CRIS	1,7	3,4
C1_CRIS	1,9	3,8
C2_CRIS	1,2	2,4
Bz(b)FLUOR	<0,5	-
Bz(k)FLUOR	nd	-
Bz(e)PIR	<0,5	-
Bz(a)PIR	<0,5	-
Perileno	nd	-
Indeno(123-cd)PIR	nd	-
DBz(ah)ANT	nd	-
Bz(ghi)PERIL	nd	-
p-TERPHd14	90,44	-
16 HPA	11314,3	-
$\Sigma$ HPA	217282,4	-

Anexo 13 Concentração dos hidrocarbonetos policíclicos aromáticos individuais em amostras de hepatopâncreas após 96 horas do bioensaio 2 (ng.g-1)

<b>Amostra</b>	<b>Controle 96h</b>	<b>Sedimento 96h</b>
NAF	1,78	nd
1ME_NAF	4,25	6,42
2Me_NAF	1,12	4,81
C2_NAF	nd	nd
C3_NAF	nd	nd
C4_NAF	nd	nd
ACENAFTY	3,04	4,30
ACE	1,43	1,58
FLUOR	8,25	5,25
C1_FLUOR	nd	2,54
C2_FLUOR	nd	nd
C3_FLUOR	nd	nd
DBZTIOF	nd	18,71
C1_DBZTIOF	nd	nd
C2_DBZTIOF	nd	nd
C3_DBZTIOF	nd	nd
FEN	nd	nd
C1_FEN	nd	50,44
C2_FEN	nd	4,68
C3_FEN	nd	7,17
C4_FEN	nd	2,15
ANT	3,18	3,30
FLUORAN	nd	40,65
PIR	nd	nd
C1_PIR	nd	1,58
C2_PIR	nd	5,22
Bz(a)ANT	nd	nd
CRIS	nd	nd
C1_CRIS	nd	nd
C2_CRIS	nd	3,47
Bz(b)FLUOR	nd	8,61
Bz(k)FLUOR	nd	nd
Bz(e)PIR	nd	nd
Bz(a)PIR	nd	nd
Perileno	nd	11,40
Indeno(123-cd)PIR	nd	nd
DBz(ah)ANT	nd	nd
Bz(ghi)PERIL	3,23	nd
p-TERPHd14	106,33	106,52
16 HPA	20,91	63,69
$\Sigma$ HPA	26,28	182,25

Anexo 14. Concentração dos hidrocarbonetos policíclicos aromáticos individuais em amostras de sedimento antes do início do e após 96 horas do bioensaio 2 (ng.g-1)

<b>Amostra</b>	<b>Início bioensaio</b>	<b>Final bioensaio</b>
NAF	< 0,23	2,09
1ME_NAF	< 0,23	0,72
2Me_NAF	< 0,23	0,78
C2_NAF	1,45	8,03
C3_NAF	5,43	10,44
C4_NAF	31,11	12,83
ACENAFTY	0,40	0,61
ACE	< 0,23	1,12
FLUOR	0,56	9,35
C1_FLUOR	5,70	33,97
C2_FLUOR	32,61	27,26
C3_FLUOR	88,15	32,31
DBZTIOF	2,51	1,69
C1_DBZTIOF	13,77	4,47
C2_DBZTIOF	72,84	9,76
C3_DBZTIOF	144,91	13,08
FEN	2,50	33,69
C1_FEN	18,47	22,32
C2_FEN	69,96	30,37
C3_FEN	146,48	87,61
C4_FEN	161,61	26,49
ANT	2,73	3,57
FLUORAN	5,98	11,60
PIR	45,55	22,55
C1_PIR	146,47	23,01
C2_PIR	251,25	34,42
Bz(a)ANT	11,20	4,44
CRIS	37,93	6,90
C1_CRIS	91,73	14,34
C2_CRIS	111,71	9,75
Bz(b)FLUOR	13,85	11,80
Bz(k)FLUOR	2,77	2,26
Bz(e)PIR	23,54	3,37
Bz(a)PIR	14,47	5,00
Perileno	10,38	5,08
Indeno(123-cd)PIR	5,04	4,58
DBz(ah)ANT	4,80	2,51
Bz(ghi)PERIL	7,06	2,72
p-TERPHd14	110,65	108,42
16 HPA	154,8	124,80
$\Sigma$ HPA	1584,9	536,90

Anexo 15. Concentração dos hidrocarbonetos policíclicos aromáticos individuais em amostras de hepatopâncreas do bioensaio 3 (ng.g-1)

<b>Amostra</b>	<b>Cont24hs</b>	<b>Cont48hs</b>	<b>Fen24hs01</b>	<b>Fen48hs</b>	<b>Fen72hs</b>
NAF	67,41	125,21	51,40	18,83	48,84
1ME_NAF	23,59	42,94	58,65	9,11	31,10
2Me_NAF	12,62	54,05	37,34	12,58	32,06
C2_NAF	<LD	<LD	<LD	<LD	<LD
C3_NAF	<LQ	<LQ	<LQ	<LQ	<LQ
C4_NAF	<LQ	<LQ	<LQ	<LQ	<LQ
ACENAFTY	<LD	5,27	27,43	6,88	16,67
ACE	22,84	2,62	15,58	1,58	10,83
FLUOR	19,22	19,16	106,85	52,27	12,84
C1_FLUOR	<LQ	<LQ	<LQ	<LQ	<LQ
C2_FLUOR	<LQ	<LQ	<LQ	<LQ	<LQ
C3_FLUOR	<LQ	<LQ	<LQ	<LQ	<LQ
DBZTIOF	14,50	236,61	1487,73	602,97	9,07
C1_DBZTIOF	<LQ	<LQ	<LQ	<LQ	<LQ
C2_DBZTIOF	<LQ	<LQ	<LQ	<LQ	<LQ
C3_DBZTIOF	<LQ	<LQ	<LQ	<LQ	<LQ
FEN	142,78	<LQ	249,83	112,80	93,01
C1_FEN	<LQ	153,68	<LQ	<LQ	<LQ
C2_FEN	<LQ	<LQ	<LQ	<LQ	<LQ
C3_FEN	<LQ	<LQ	<LQ	<LQ	<LQ
C4_FEN	<LQ	<LQ	<LQ	<LQ	<LQ
ANT	<LD	<LD	<LD	<LD	<LD
FLUORAN	<LD	<LD	<LD	<LD	<LD
PIR	<LD	<LD	22,26	107,23	34,57
C1_PIR	<LQ	<LQ	<LQ	<LQ	<LQ
C2_PIR	<LQ	<LQ	<LQ	<LQ	<LQ
Bz(a)ANT	<LD	<LD	<LD	<LD	<LD
CRIS	<LD	<LD	<LD	<LD	<LD
C1_CRIS	<LQ	<LQ	<LQ	<LQ	<LQ
C2_CRIS	<LQ	<LQ	<LQ	<LQ	<LQ
Bz(b)FLUOR	<LD	<LD	<LD	<LD	<LD
Bz(k)FLUOR	<LD	<LD	<LD	<LD	<LD
Bz(e)PIR	<LD	<LD	<LD	<LD	<LD
Bz(a)PIR	<LD	<LD	<LD	37,89	<LD
Perileno	<LD	<LD	<LD	<LD	<LD
Indeno(123-cd)PIR	<LD	<LD	<LD	<LD	<LD
DBz(ah)ANT	<LD	<LD	<LD	<LD	<LD
Bz(ghi)PERIL	<LD	<LD	<LD	<LD	<LD
16 HPA	252,25	152,26	10866,52	4172,76	216,76
$\Sigma$ HPA	302,97	639,55	15485,26	7828,76	19167,67
p-TERPHd14	75,53	73,69	66,41	74,36	65,34

Anexo 16. Concentração dos hidrocarbonetos policíclicos aromáticos individuais em amostras de hepatopâncreas inoculadas com 1-Metil fenantreno e 2,6,9-Trimetil fenantreno do bioensaio 3 (ng.g<sup>-1</sup>)

Amostra	1Mef24hs	1Mef48hs	1Mef72hs	Trimef24hs	Trimef48hs	Trimef72hs
NAF	40,46	57,31	587,02	2032,27	30,58	58,67
1ME_NAF	49,28	53,47	529,46	877,56	9,06	37,37
2Me_NAF	47,14	51,74	626,99	442,11	27,15	36,05
C2_NAF	<LD	<LD	128,19	<LD	<LD	<LD
C3_NAF	<LQ	<LQ	60,70	<LQ	<LQ	<LQ
C4_NAF	<LQ	<LQ	91,10	<LQ	<LQ	<LQ
ACENAFTY	18,09	<LD	115,81	36,48	12,58	14,12
ACE	25,66	<LD	88,65	<LD	<LD	<LD
FLUOR	16,40	<LD	75,65	<LD	11,71	9,75
C1_FLUOR	<LQ	<LQ	51,03	<LQ	<LQ	<LQ
C2_FLUOR	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
C3_FLUOR	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
DBZTIOF	21,45	9,86	0,87	<LD	<LD	19,64
C1_DBZTIOF	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
C2_DBZTIOF	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
C3_DBZTIOF	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
FEN	<LQ	4,80	3,37	<LQ	<LQ	3,45
C1_FEN	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
C2_FEN	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
C3_FEN	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
C4_FEN	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
ANT	133,62	31,09	10,27	47,41	138,18	165,99
FLUORAN	11,30	86,54	<LD	<LD	<LD	<LD
PIR	82,89	125,71	0,87	<LD	<LD	<LD
C1_PIR	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
C2_PIR	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
Bz(a)ANT	<LD	<LD	<LD	<LD	<LD	<LD
CRIS	<LD	<LD	0,96	<LD	<LD	<LD
C1_CRIS	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
C2_CRIS	<LQ	<LQ	<LQ	<LQ	<LQ	<LQ
Bz(b)FLUOR	<LD	9,69	<LD	<LD	<LD	<LD
Bz(k)FLUOR	<LD	7,17	<LD	<LD	<LD	<LD
Bz(e)PIR	<LD	<LD	<LD	<LD	<LD	<LD
Bz(a)PIR	<LD	<LD	<LD	<LD	<LD	<LD
Perileno	161,48	29,28	<LD	<LD	<LD	<LD
Indeno(123-cd)PIR	<LD	<LD	<LD	<LD	<LD	<LD
DBz(ah)ANT	<LD	<LD	<LD	<LD	<LD	<LD
Bz(ghi)PERIL	<LD	<LD	<LD	<LD	<LD	<LD
16 HPA	379,10	485,45	997,12	2141,67	246,58	369,17
$\Sigma$ HPA	658,45	3262,12	2485,45	3461,34	15511,49	2863,72
p-TERPHd14	66,53	95,18	74,80	71,57	64,26	69,32